



Enzymatic Activities of Actinomycetes from the Soil Samples of Jammu and Kashmir

Shawkat Ahmad Thokroo
Department of Microbial Biotechnology
IIIM Jammu, India

Abstract:

The present study focused on isolation of actinomycetes from the soil samples of jammu and Kashmir can provide rich source of enzyme producing actinomycetes. Totally, 52 different actinomycetes were isolated from the soil samples of jammu and Kashmir. 90 actinomycetal isolates were identified up to generic level. It was found that these actinomycetal isolates were belonging to *Streptomyces*, *Micromonospora*, *Intrasporangium*, *Saccharopolyspora*, *Streptosporangium*, *Rhodococcus*, *Saccharomonospora* and *Nocardia*. It has been found that out of 40 actinomycetes, 32 (80%), 27 (67.50%), 34 (85%) and 25 (62.5%) number of actinomycetes possessing amylase, protease, lipase and cellulose activity respectively.

Keywords: actinomycetes, amylase, protease, lipase, cellulose, oil samples.

I. INTRODUCTION

The name “Actinomycetes” was derived from Greek words ‘aktis’ (a ray) and ‘mykes’ (fungus). Actinomycetes are true bacteria (related to Corynebacteria and mycobacteria) but they form long branching filaments that resembles the hyphae of fungi, yet possess sufficient distinctive features to classify them into a separate actinomycete. Actinomycetes are best known for their ability to produce antibiotics and are gram positive bacteria which comprise a group of branching unicellular microorganisms.

II. MATERIAL AND METHODS

Take 250 ml of CSA broth was put it in 1 litre of distilled water. PH was adjusted to 7 and then 2% agar was added. The medium was autoclaved at 121°C, 15 pounds/inch for 15 minutes.

Sterility testing of Medium

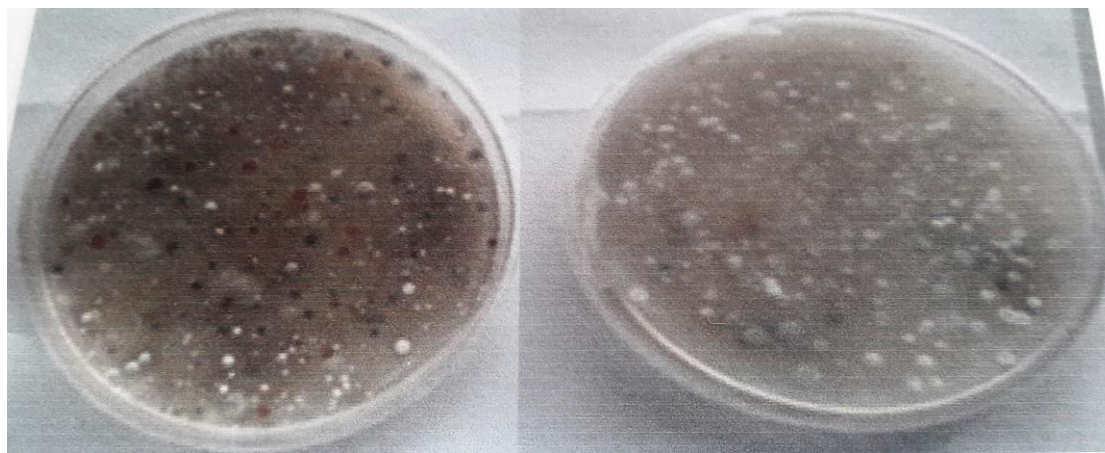
The autoclaved media were incubated at 37°C for overnight for sterility testing.

Isolation of actinomycetes strain from soil samples of Jammu and Kashmir

CSA media was prepared, sterilized and cooled. The media was poured on the Petri plates and incubated for overnight at 37° C to check sterility. Prepared three distilled water test tubes (9 ml) each and autoclaved them. With the help of sterilized spatula, added 1g of soil samples in sterilized distilled water tube and mixed it and then from it made dilutions up to 10⁻³. By using micropipette and sterilized tips 100 µl of inoculum was poured on CSA- plates and spread it with sterilized and cooled glass spreader. Incubated the plates at 28° C for few days and observed for the appearance of colonies. Picked up the colonies appeared different in color with the help of sterilized tooth picks and incubated on CSA plates at 28°C.

Purification of isolated colonies

Prepare CSA media sterilized and cooled. Poured the Petri plates and incubated for overnight at 37 C to check sterility. Then streak the inoculum from master plates (cyps). Incubate Petri plates at 28°C for 5days. After 5th day growth appears on plates.



III. ENZYMATIC ACTIVITY OF ACTINOMYCETES

The actinomycetes isolates were inoculated on suitable medium by streak or spot inoculation method in order to check different enzymatic degradative activities. For amylase activity, casein starch agar with 1% starch was prepared. The 10 day old grown culture plugs were spot inoculated on to the medium and incubated at 28° C for 72 hours. The gram's iodine stain was spread on plate and left for 5 minutes. The organism which secretes amylase, produce zone of clearance around the growth. For protease activity, casein starch agar with 1% skimmed milk (SM) was prepared. The 10 day old grown culture plugs were spot inoculated on to the medium and incubated at 28° C for 72 hours. The organism which secretes protease, produce zone of clearance around the growth. For lipase activity, in one flask

take 98ml 50mM Tris Hcl, pH 6.8, and 1.2 gm agar autoclave it. In another flask take 2ml Tris Hcl and autoclave it and add 300µ l of tributyrin (TB) and 30µ l of tween 20 after autoclaving and then mix both flasks and pour on the plates. The 10 day old grown culture plugs were spot inoculated on to the medium and incubate at 28° C for 72 hours. The organism which secretes lipase, produces zone of clearance around the growth.

For cellulose activity, casein starch agar with 1% carboxymethyl cellulose (CMC) was prepared. The 10 days old grown culture plugs were spot inoculated on to the medium and incubated at 28° C for 72 hours. The congo red stain (2%) was spread on plate and left for 5 minutes. Plates were destained with 1M NaCl. The organism which secretes cellulose, produce zone of clearance around the growth.

Table.1. Studies on the Enzymatic Activities on Actinomycetes

Sr. No	Enzyme	Medium	Incubation period (days)	Criteria of positive activity
1	Amylase	CSA with 1% starch	7	Zone of clearance
2	Protease	CSA with 1% skimmed milk	7	Zone of clearance
3	Lipase	98 ml of 50 mM Tris Hcl+ 1.2 gm in 1 flask & 2ml Tris+ 300 µl Tributyrin+30 µl Tween 20	7	Zone of clearance
4	Cellulose	CSA with 1% CMC	7	Zone of clearance

IV. RESULT AND DISCUSSION:

The enzymatic activity of actinomycetes is shown above and only few culture shown good enzymatic activity. Those culture which shown good zone of clearance will show good enzymatic activity. Actinomycetes have been a source of a numerous useful products including pharmacerticals, agrochemicals, enzymes for use in a number of industrial applications from food industry to paper making.

V. REFERENCE

[1]. Hagerdal, B. Ferchak, J.D. and Pye, E.K. (1980). saccharification of cellulose by the cellulolytic enzyme

[2]. system of *Thermomonopora* Species I. Stability of cellulolytic activities with respect to time, temperature and PH. Biotechnology and Bio engineering. 22 : 1515-1526. Holt, J. G. (1994). Bergey's Manual of Determinative

[3]. Bacteriology 9th edition (Willian and Wilkin Baltimore), pp 667-669. Kawato, N. and Shinobu, R. (1959). On *Streptomyces herbaricolor*, nov SP supplement: a simple technique for the microscopic observation. Mem. Saka Univ. Lib. Arts Educ. J.Nat. Sci. 8 : 114-119. Konde, B.K. (1978).

[4]. Studies on soil *Streptomyces* from Maharashtra Ph.D. (Argi) Thesis. Uni. Poona, Pune. Lee, Y.H. and Fan, H.T. (1981). Properties and mode of action of cellulases. In: products from alkanes, cellulose and other Feed stock's 9A. Fiechter. ed,

[5]. Moreira, A.R., Philips, J.A. and Humprey, A.E. (1981). Production of cellulose by *Thermonospora* Species. Biotechnology and Bioengineering. 23: 1339- 1347.

[6]. Navine, B. Ghanem, Soraya, A., Sobry, Zeinab M.El.Sherif , and Gehan A., Abu El-Elq (2000). Isolation and enumeration of marine actinomycetes from sea water and sediments in Alexandria. J. Gen.Appl. Microbio. 46: 105-111.

[7]. Nawani, N.N.; Kapdnis, B.P. Das, A.S., Rao, A.D. and Mahajon, S.K. (2002). Purification and characterization of a thermophilic and acidophilic chitinase from *Microbiospora* sp V2. J. Appl. Microbiol. 93: 9645-975.

[8]. Obi, S.K.C. and Obido, F.J.C. (1984). Some properties of a highly thermostable – amylase from *Thermoactinomyces* Sp. Candian J. of Microbiology. 30: 780-785.

[9]. *Thermonospora fusca* strain 190 T H. Can .J. Microbio. 21:1842-1848

[10]. Dhanasekaran, D., Rajakumar, G.,Sivamani, Selvamani, S., Paneerselvam, A. and Thajuddin, N. (2005). Screening of salt pan actinomycetes for antimicrobial agents .Internet J. Microbiol. 8:1-8.

[11]. Fogarty, W.M. (1983). In: Microbial Enzymes and Biotechnology (W.M. Fogarty. Ed), Applied Science Publishers, London, pp 1 – 29.

[12].Okawa, Y&Yamaguchi, T. (1975). Studies on Phospholipase from *Streptomyces* III Purification and properties of *Streptomyces habjioensis* Phospholipase C. J. of Biochemistry. 78 : 537-545.

[13]. Patke, D.S. and Dey, S.(1996). Characterization of enzyme and genetic studies of thermophilic *Streptomyces*. Ph.D. Thesis. University of Pune, Pune. Ratnakala, R. and Chandrika,

- V. (1993). Effect of different media for isolation, growth and maintenance of actinomycetes from mangrove sediments. *Indian J. Mar Sci.* 22: 297-299.
- [14]. Shejul, M.S. (1999). Studies on heterotrophic filamentous prokaryotes from aquatic habitats. Ph.D. Thesis, University of Pune, Pune. Stanford, T.L., Stamford, N.P. Coelho, L.C and Araujo, J.M. (2001).
- [15]. Production and characterization of a thermostable alpha-amylase from *Nocardiopsis SP* endophyte of yambean. *Bioresour Technol.* 76: 137-141.
- [16]. Stapp, C. (1953). Untersuchungen uber Aktinomyzeten des Bodens Zentr. Backteriol. Parasitenk. Abt.II. 107: 129 – 150.
- [17]. Stutzenberger, F.J. (1972). Cellulase production by *Thermomonospora curvata* isolated from municipal solid waste compost. *App. Microbiol.* 22: 147.
- [18]. Surajit Das, Lyla, P.S. and Ajmal Khan (2006). Marine Microbial diversity and ecology importance and future perspectives. *Curr. Sci.* 90:1325-1334.
- [20]. Williams, S.T., Goodfellow, M., Anderson, G., Willington, E.M.H. Sneath, P.H.A. and Sackin, M. J. (1983).
- [21]. Numerical classification of *Streptomyces* and related genera. *Journal of General Microbiology.* 129: 1743-1813
- [22]. Saadoun, I., Rawashdeh, R., Dayeh, T., Ababneh, Q., Mahasneh, A. 2007. Isolation, characterization and screening for fiber hydrolytic enzymesproducing streptomycetes of Jordanian forest soils. *Biotechnology*, 6(1): 120 128.
- [23]. Salahuddin, K., Prasad, R., Kumar, S., Visavadia Manish, D. 2011. Isolation of soil thermophilic strains of actinomycetes for the production of - amylase. *Afr. J. Biotechnol.*, 10(77): 17831 17836.
- [24]. Santos, E.O., Martins, M.L.L. 2003. Effect of the medium composition on formation of amylase by *Bacillus* sp. *Braz. Arch. Biol. Technol.*, 46(1): 129 134.
- [25]. Sharma, R., Chisti, Y., Banerjee, U.C. 2001. Production, purification, characterization, and applications of lipases. *Biotechnol. Adv.*, 19: 627 662.
- [26]. Shaw, F.J., Ou-Lee, T.M. 1984. Studies on the-amylase from the germinated rice seeds. *Bot. Bull. Acad. Sin.*, 23: 41 46. Shirling, E.B., Gottlieb, D. 1966. Methods for characterization of *Streptomyces* species. *Int. J. Syst. Bacteriol.*, 18(3):313 340.
- [27]. Shori Ghadeer, B.O., Mohamed Sonya, H., Abdel-Salam Shima, M., Sadik, A.S.
2012. Characterization of streptomycetes having antibiosis activities isolated from soil in westernregion of KSA. *Pak. J. Biotechnol.*, 9: 21 38.
- [28]. Soares, A.C.F., Sousa, C. da S., Garrido, M. da S., Perez, J.O., de Almeida, N.S. 2006. Soil streptomycetes with *in vitro* activity against the yam pathogens *Curvularia eragrostides* and *Colletotrichum gloeosporioides*. *Braz. J. Microbiol.*, 37(4): 456-461.
- [29]. Stefka, A.N., Nikoleta, T., Ljubomira, Y. 2004. Taxonomy of *Streptomyces* sp. strain 3B. *J. Cult Collect.*, 4: 36 42.
- [30]. Sztajer, H., Maliszewska, I., Wieczorek, J. 1988. Production of exogenous lipases by bacteria, fungi, and actinomycetes. *Enzyme Microb. Technol.*, 10(8): 492 497.
- [31]. Techapun, C., Poosaran, N., Watanabe, M., Sasaki, K. 2003. Thermostable and alkaline-tolerant microbial cellulasefree xylanases produced from agricultural wastes and the properties required for use in pulp bleaching bioprocesses: a review. *Proc. Biochem.*, 38(9): 1327 1340.
- [32]. Techapun, C., Sinsuwongwat, S., Watanabe, Sasaki, Poosaran, N. 2002. Production of cellulase-free xylanase by a thermotolerant *Streptomyces* sp. grown on agricultural waste and media optimization using mixture design and Plackett Burman experimental design methods. *Biotechnol. Lett.*, 24(17): 1437 1442.
- [33]. Tandler, M.D., Burkholder, P.R. 1961. Studies on the thermophilic actinomycetes I. methods of cultivation. *Appl. Environ. Microbiol.*, 9(5): 394 399.
- [34]. Waksman, S.A., Lechevalier, H.A. 1961. The actinomycetes, Vol. II. Classification, identification and descriptions of genera and species. The Williams and Wilkins, Co. Baltimore, USA.
- [35]. Yang, S.S., Wang, J.Y. 1999. Protease and amylase production of *Streptomyces rimosus* in submerged and solid state cultivations. *Bot. Bull. Acad. Sin.*, 40: 259 265.