



Effects of Heavy Metals and Physicochemical Parameters on the Aquatic Biota of Elechi Creek, Port Harcourt

Dagde Kenneth K¹, Iregbu Precious O¹, Nwideduh Godson N³
Rivers State University, Nkpolu-Oroworukwo, Port Harcourt

Abstract:

Accumulation of heavy metals chromium (Cr) and lead (Pb) in water, sediment and plankton of Elechi creek were studied for six (6) months covering three months of dry and wet seasons in three different locations (stations). Physicochemical parameters were studied and were found to correlate with heavy metals. Experimental and mathematical modelling methods were adopted in this research for the determination of the heavy metals concentration. The mathematical model was developed based on the principle of statistical modelling using regression analysis. The results of the concentration of heavy metals from the model were compared with the experimental results for each metal in water, sediment and plankton and the results showed reasonable agreement with slight deviations as follows: for Chromium (Cr) -0.17 to 0.15, -0.12 to 0.13, and -0.04 to 0.06, for lead (Pb) -0.29 to 0.1, -0.37 to 0.21, and -0.05 to 0.18, in water, sediment and plankton respectively. Concentration of heavy metal in plankton, sediment and water varied seasonally except lead in water and sediment. The results indicated that changing water quality of Elechi creek affected the aquatic biota in terms of diversity and abundance and so the mathematical tools can be used to monitor the ecological status of the creek.

Keywords: Heavy metals, Plankton and Physicochemical parameters.

1.0 INTRODUCTION

Heavy metals are inorganic elements essential for plant growth, in trace or very minute quantities, toxic and poisonous in relatively higher concentrations, biologically non-degradable but easily assailable and bio-accumulated in the protoplasm of aquatic organisms. They are also considered to be metals whose densities are greater than 5gcm^{-3} . In natural aquatic ecosystems, metals occur in low concentration in monogram to microgram per litre. Examples of these heavy metals are lead (Pb), mercury (Hg), zinc (Zn), cadmium (Cd), iron (Fe), copper (Cu) etc. The occurrence of these metals contaminants especially in excess of natural levels in recent times has become a problem of increasing concern. This is because its accumulation in the sediments, water and tissues of aquatic micro biota (plants and animals) affect their productivity (Yan *et al.*, 2007). The accumulation of heavy metals in the sediments and water affect the physicochemical properties of the aquatic environment and so alter the entire physiological and metabolic processes of the aquatic biota. He *et al.* (2005) disclosed that in the aquatic ecosystems, heavy metals have received considerable attention due to their toxicity and accumulation in biota. Aquatic organisms have been used in comparative monitoring of pollution effects in different systems and to locate sources of toxicants (Duffus, 2002). Bio-monitoring approach has proved to promising as a reliable means of quantifying biological effects of complex effluents (Bradl, 2002). Although a large number of aquatic organisms have been used for assessment, many other researchers have recognized the importance of algae as indicator in assessment and evaluation of pollution (Arruti *et al.*, 2010). Lake ecosystems are increasingly affected by various anthropogenic impacts, such as excess of nutrients causing eutrophication, toxic contamination of industrial, agricultural and domestic origin, as well as heat pollution reaching the lakes through their catchments area and the atmosphere. Typical results of human activities proved to be

the cause of elevated levels of heavy metals present in fresh water, and among these microelements lead (Pb), cadmium (Cd), mercury (Hg), chromium (Cr) are most specific (Farkaset *al.*, 2001). They are considered to be one of the most important pollutants of the aquatic ecosystems due to their environmental persistence and tendency to be concentrated in aquatic organisms (Stern, 2010). In addition, heavy metals show harmful effects even at very low concentration on the aquatic organisms including plankton, aquatic plants, invertebrates and vertebrates. This study will be conducted to determine the accumulation of heavy metals in water and plankton (phytoplankton and zooplankton) of Elechi Creek. Physicochemical parameters refer to the characteristics of water affecting the survival, growth, reproduction, production and general management of aquatic organisms (Zhang, *et al.*, 2011). Researches disclosed that maintaining optimum water quality means keeping the water at an optimum for the physiological requirement of aquatic species, precisely fish and plankton. Bradl, (2002) also disclosed that physicochemical parameters refers to all physical, biological and chemical variables affecting the desirability of water for any particular use. These variables therefore include temperature, salinity, alkalinity, dissolved oxygen, biological oxygen demand, total hydrocarbon etc.

2.0 MATERIALS AND METHODS

2.1 Water Sampling and Analysis

Water samples were collected from the water surface at the various sampling stations using one-litre plastic containers Merrett, (2000). The water samples collected were kept in an acid rinsed polyethylene bottles and later filtered through the $0.45\mu\text{m}$ membrane filter and acidified with concentrated nitric acid to a pH of 2 Tahir *et al.*, (2008). The acidified samples were kept for determination of heavy metals using Buck Scientific Atomic Absorption/Emission Spectrophotometry

200A. It was also used for the determination of some physicochemical parameters of water.

2.2 Sediment Sampling and Analysis

About 4-5cm of the top of sediment samples was collected from each of the three stations which are approximately 500m apart using grab sampler. At least 100g of the sediment samples was collected in glass containers as recommended by ASTM (1990) Guide for collection, storage, and characterization for toxicological testing and transported in ice chest at 4°C to the laboratory for preparation, treatment and analysis. In the laboratory, representative portions were oven dried at 200°C to constant weight, crushed and passed through a 2mm mesh sieve to do away with coarse materials. Later, about 2g of the sample was weighed with the triple beam balance and turned into the evaporating dish. 25ml of distilled water was added followed by 0.5ml of nitric acid (HNO₃) and 5ml of hydrochloric acid (HCl) and thoroughly digested for about 45m on the steam bath to release the total metal content from the sediment into solution (Loring and Rantala, 1992, APHA, 1985). The samples were extracted by filtering with Whatman No. 1 filter paper into 50ml volumetric flasks and the filtrate diluted with distilled water to the 50ml mark and poured into the vial bottle which was kept in the freezer until it was sent to the laboratory for determination of heavy metals using Buck Scientific Atomic Absorption/Emission Spectrophotometry 200A.

2.3 Biota Sampling and Analysis

Biota (plankton) samples were collected by throwing plankton net (25 µm) five minutes duration. Samples were collected by horizontal dragging of the net in water using speed boat in each of the station at 500m apart for 10 minutes. Collected samples were transferred into plastic samples bottles and were preserved in 4% formalin solution (Ezra and Nwankwo, 2001). The plankton samples collected were also used for heavy metal determination. The samples were preserved for some time and later sent to the laboratory for enumeration and identification.

2.4 Laboratory Methods

2.4.1 Plankton Enumeration and Identification

The concentrated plankton sample were thoroughly mixed and 1ml of the sample was drawn with a dropping pipette into a Sedgwick Kafer counting chamber and was examined using an inverted microscope (X100) or the electrical Nikon biological microscope or digital microscope. Plankton were identified and the total number per species were recorded using keys and checklist of Moss (2010), Thorp and Covich (1991). The samples collected for plankton analysis were filtered through ash-less filter paper with concentrated plankton left in laboratory for some time for small evaporation of water. Age-weight was determined following Tahir *et al.*; (2008). Material was then dried in oven at 90°C for 24 hours and its dry weight was noted. The material was then kept at 550°C for 4 hours and then cooled down. It was digested at 80°C by adding 20 ml of 1.5N HCl, filtered into 100 ml test tubes, 100 ml distilled water was added and samples were analysed with a

For chromium

$$Scr_{rw} = \left(\frac{n \sum st - (\sum s)(\sum t)}{n \sum t^2 - (\sum t)^2} \right)^{t+} \left(\frac{\sum s}{n} - \frac{n \sum st - (\sum s)(\sum t)}{n(\sum t^2) - (\sum t)^2} \cdot \frac{\sum t}{n} \right)_{crw} \quad (1)$$

$$Scr_{rs} = \left(\frac{n \sum st - (\sum s)(\sum t)}{n \sum t^2 - (\sum t)^2} \right)^{t+} \left(\frac{\sum s}{n} - \frac{n \sum st - (\sum s)(\sum t)}{n(\sum t^2) - (\sum t)^2} \cdot \frac{\sum t}{n} \right)_{crs} \quad (2)$$

Hitachi Z 8200 polarized Zeeman AAS (Atomic Absorption Spectrophotometer) at the Institute of Pollution Studies, Rivers State University Port Harcourt and soil science laboratory respectively.

2.4.2 Analysis of Data

The estimated count of plankton was observed per litre and subjected to the formula modified by Bradl, (2002).

$$\text{Density or plankton (ind/ml)} = T \left(\frac{100 \times \text{Volume of concentration}}{AN \times \text{volume of sample}} \right)^J$$

where, T = total number of plankton counted, A = Area of grid in mm², i- number of grids employed, 100 = area of counting chamber in mm (Tang *et al.*, 2010).

2.5 Physicochemical Parameters

The various water parameters were studied following the normal procedures as recorded in the standard method.

2.5.1 Temperature

The water temperate was measured using (in-situ) mercury in bulbs thermometers at the various stations.

2.5.2 pH

The pH values of the water and sediment samples were determined in the laboratory using an EIL Model 720 pH meter or model EC10 each. The pH was determined by simply dipping the electrode into a 200ml of water that had been stirred and the reading was subsequently read off the meter.

2.5.3 Dissolved Oxygen

Water samples were collected from each station with 70ml DO bottles followed by the addition of each Winkler Solution I (manganese II tetraoxosulphate IV monohydrate and II (sodium hydroxide and sodium iodide) to fix it in the sample. Also, 0.5ml of concentrated tetraoxosulphate VI acid (H₂SO₄) was added to liberate the iodine equivalent of dissolved oxygen in the titration with the thiosulphate.

2.5.4 Electrical Conductivity

This will be determined from the water sample using Griffin Conductance Bridge. Electrical conductivity will be measured or expressed as µs/cm.

2.5.5 Biological Oxygen Demand (BOD)

Samples for this variable were collected from the respective stations with 70ml bottles. The samples were stored 5 days before fixing and titration. After this Winkler I and II solutions were added to the samples followed by 0.5ml of concentrated H₂SO₄. The bottles were inverted after which 25ml of the samples were transferred into 100ml flask and 2 drops of starch were added.

2.6 Mathematical Models

Using regression analysis as a statistical modelling approach in the determination of heavy metal accumulation in water, sediment and plankton, the following representative models for chromium, lead, zinc and copper were obtained as:

$$Scrp = \left(\frac{n \sum st - (\sum s)(\sum t)}{n \sum t^2 - (\sum t)^2} \right)^{r+} \left(\frac{\sum s}{n} - \frac{n \sum st - (\sum s)(\sum t)}{n(\sum t^2) - (\sum t)^2} \frac{\sum t}{n} \right)_{crp} \quad (3)$$

For lead

$$Spbw = \left(\frac{n \sum st - (\sum s)(\sum t)}{n \sum t^2 - (\sum t)^2} \right)^{t+} \left(\frac{\sum s}{n} - \frac{n \sum st - (\sum s)(\sum t)}{n(\sum t^2) - (\sum t)^2} \frac{\sum t}{n} \right)_{pbw} \quad (4)$$

$$Spbs = \left(\frac{n \sum st - (\sum s)(\sum t)}{n \sum t^2 - (\sum t)^2} \right)^{t+} \left(\frac{\sum s}{n} - \frac{n \sum st - (\sum s)(\sum t)}{n(\sum t^2) - (\sum t)^2} \frac{\sum t}{n} \right)_{pbs} \quad (5)$$

$$Spbp = \left(\frac{n \sum st - (\sum s)(\sum t)}{n \sum t^2 - (\sum t)^2} \right)^{t+} \left(\frac{\sum s}{n} - \frac{n \sum st - (\sum s)(\sum t)}{n(\sum t^2) - (\sum t)^2} \frac{\sum t}{n} \right)_{pbp} \quad (6)$$

3.0 RESULTS AND DISCUSSION

3.1 Physicochemical Parameters of Water

The results of the parameters studied, namely temperature, pH, electrical conductivity, dissolved oxygen (DO), biological oxygen demand (BOD), chemical oxygen demand (COD) and total hydro-carbon compound (THC) are presented.

3.1.1 Temperature

The temperature value recorded ranged from 25 -28°C with the lowest mean (26.07 ± 0.83)⁰C observed in station 2 and the highest (26. 22 ± 1.09⁰C) observed in station 1. The lowest monthly temperature (25 .40 ± 0.10⁰C) was observed in January while the highest (27 .30 ± 0.61)⁰C was observed in September. Seasonally, temperature values were higher in the wet season (26.53 ± 0.84⁰C) than dry season (25. 73 ± 0.58⁰C) though there was no significant difference (Fig. 1 and Fig. 2).

3.1.2 pH: pH value ranged from 5.90 -7.00 with the overall mean, 6.33 ± 0.31. The lowest pH (6.18 + 0.25) was recorded in station 1 while the highest (6.50 ± 0.42) was observed in station 2. The lowest monthly pH (6.03 ± 0.06) was observed in August while the highest value (6.60 ± 0.17) was observed in February (Fig. 1 and Fig.2).

3.1.3 Electrical Conductivity(EC)

Electrical Conductivity (EC) ranged from 13200 - 24500µs/cm with the overall mean of 20.288.89 ± 3824. 363 µs/cm. The lowest spatial value (15.216, 67 ± 13. 65.89 µs/cm while the highest spatial mean (23266.67 ± 708.99 µs/cm. The lowest monthly value (19233.33 ± 5278. 66 µs/cm) was observed in January while the highest value (21400 ± 3811.82 v s/cm). EC shows significant difference with wet season value (209988.89 + 3628.57 µs/cm) higher than the dry season value (19588. 89 ±410014 µs/cm) (Fig. 1 and Fig. 2).

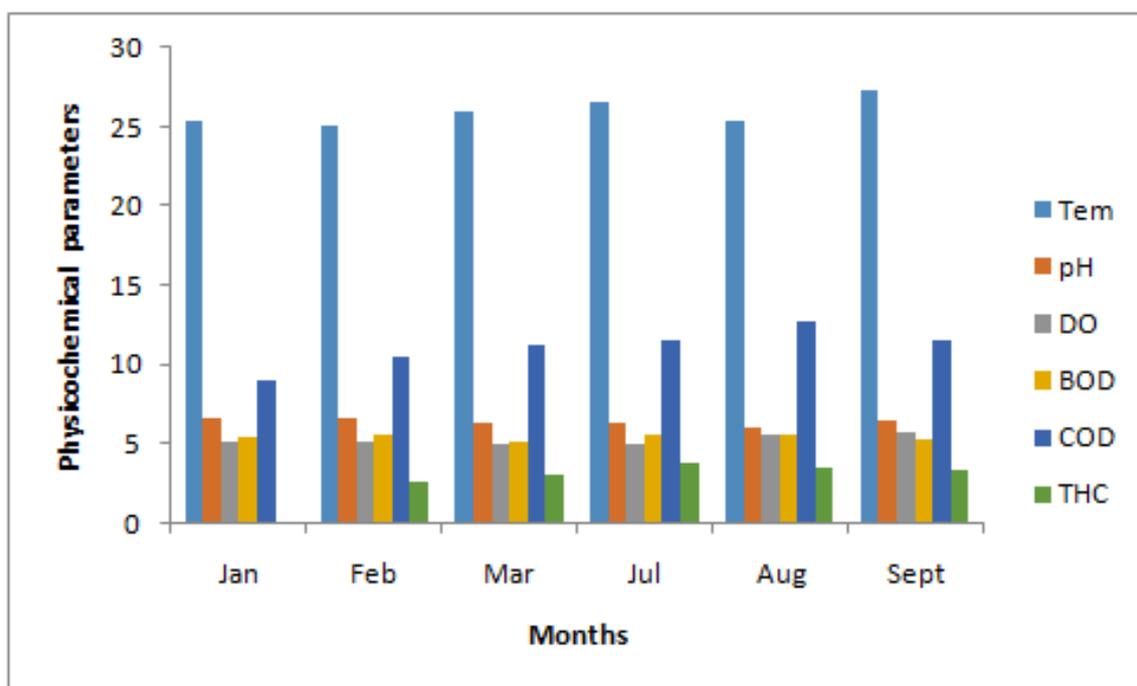


Figure.1. Mean Values of Physicochemical Parameters of Water

3.1.4 Dissolved Oxygen (DO)

The DO values, range from 3.20 - 7.3mg/l with the overall mean of 5.13 ± 12mg/l. The lowest spatial mean value (4.47 ± 1.04mg/l) was observed in station 3 while the highest value (6.37 ± 0.5 mg/l) was recorded in station 2. The lowest

monthly value (4.87 ± 1.65mg/l) was observed in March while the highest mean value 5.67± 0.58mg/l was observed in September. Seasonally, wet season value was higher than the dry season value (Fig. 1 and Fig. 2).

3.1.5 Biological Oxygen Demand (BOD):

Biological Oxygen Demand (BOD) ranged from 1.40 - 10.0mg/l with the overall mean of 6.30 ± 2.38 mg/l. Spatially, station 3 (3.43 ± 1.35 mg/l) recorded the lowest BOD while the highest value (6.98 ± 0.09 mg/l) was recorded in station 2. The

lowest monthly value (5.57 ± 2.93 mg/l) was observed in February while the highest (7.33 ± 2.52 mg/l) was recorded in station 3. Seasonally, wet season value (6.89 ± 2.21 mg/l) was higher than the dry reason value (5.71 ± 2.53 mg/l) (Fig. 1 and Fig. 2).

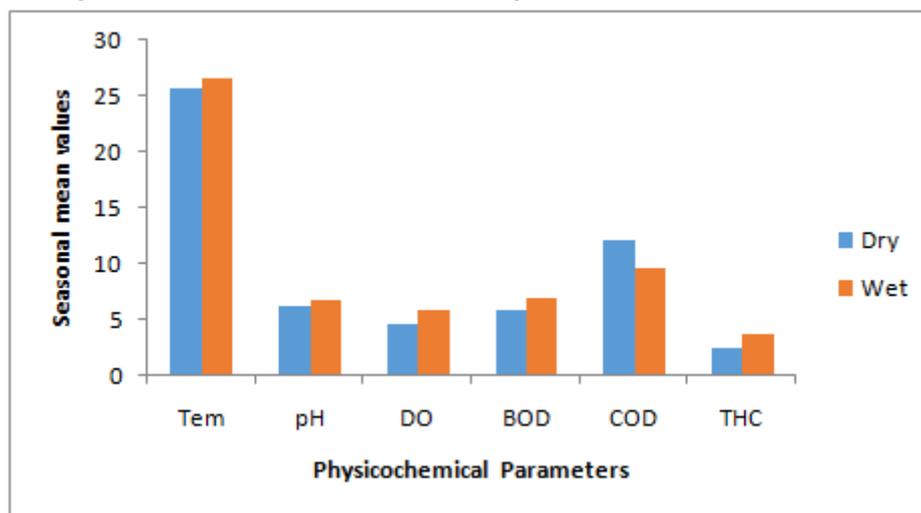


Figure.2. Seasonal Mean Values of Physicochemical Parameters in Water

3.1.6 Chemical Oxygen Demand (COD)

COD values ranged from 2.20 - 13.60mg/l with the overall mean value of 10.82 ± 3.10 mg/l. Spatially, station 3 recorded the lowest COD (8.10 ± 4.22 mg/l) while station I recorded the highest value (12.40 ± 0.85 mg/l). The lowest monthly value (9.07 ± 3.83 mg/l) was observed in January while the highest value (12.67 ± 0.58 mg/l) was observed in August. Seasonally, wet season value (12.11 ± 1.05) recorded higher value than dry season (9.53 ± 3.95 mg/l) with significant difference (Fig. 1 and Fig. 2).

3.1.7 Total Hydrocarbon Compound (THC)

Total hydrocarbon compound observed ranged from 0 -5mg/l with the overall mean value of 3.04 ± 1.79 . The lowest spatial value (1.85 ± 1.16 mg/l) was observed in station 1 while the highest value (3.63 ± 1.92 mg/l) was observed in station 2 and 3. The lowest temporal value of THC was observed in January while the highest value (4.00 ± 1.00 mg/l) was recorded in July and August. Seasonally, wet season value (3.67 ± 1.00) was higher than the dry season value (2.40 ± 2.20 mg/l) (Fig. 1 and Fig. 2).

3.2 Heavy Metals in Water

Table .1. Spatial Mean Values of Heavy Metals in Water

Metal/Station	Cr	Pb
1	0.36 ± 0.18	0.15 ± 0.12
2	0.18 ± 0.11	0.33 ± 0.13
3	0.25 ± 0.27	$0-23 \pm 0.16$
Overall mean	0.30 ± 0.21	0.20 ± 0.12

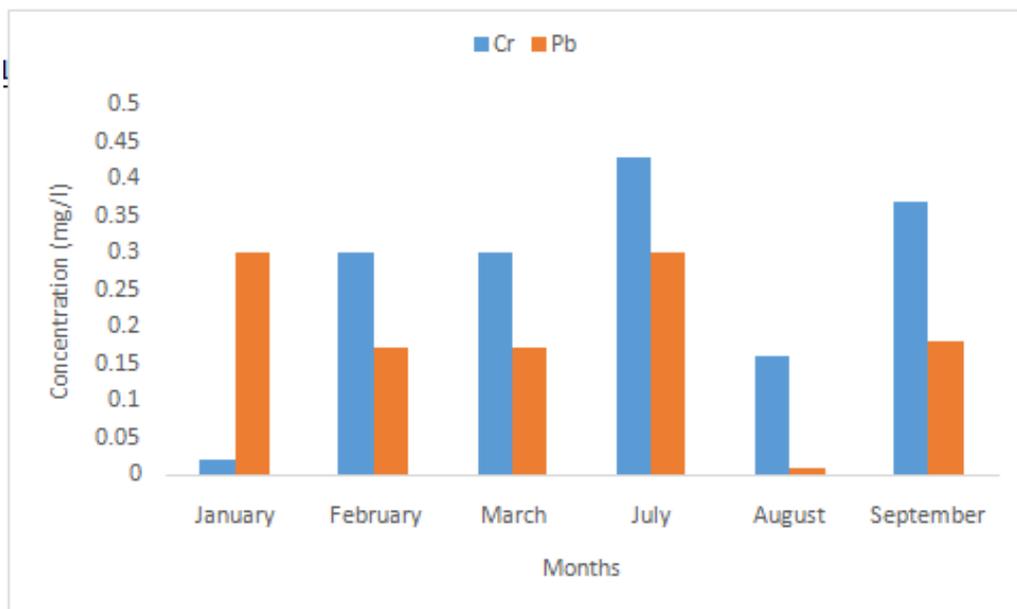


Figure.3. Mean Values of Heavy Metals in Water

Chromium is lowest in January (0.02) but recorded highest (0.43 ± 0.20) in July. Lead is lowest (0.001 ± 0.01mg/L) in

August but highest in January. In terms of station, chromium in station 2 < 3 < 1, lead in station 1 < 3 < 2

3.3. Heavy Metals in Sediment

Table.2. Spatial Mean Values of Heavy Metals in Sediment

Station	Cr	Pb
1	0.47±0.20	0.21±0.10
2	0.19±0.10	0.19±0.09
3	0.37±0.10	0.17±0.10
Overall Mean	0.37±0.10	0.17±0.10

In. terms of station, chromium in station 1 >3 >2, lead in station 3 < 2 < 1. The heavy metal concentration in the water is higher than the sediment but lower than the plankton.

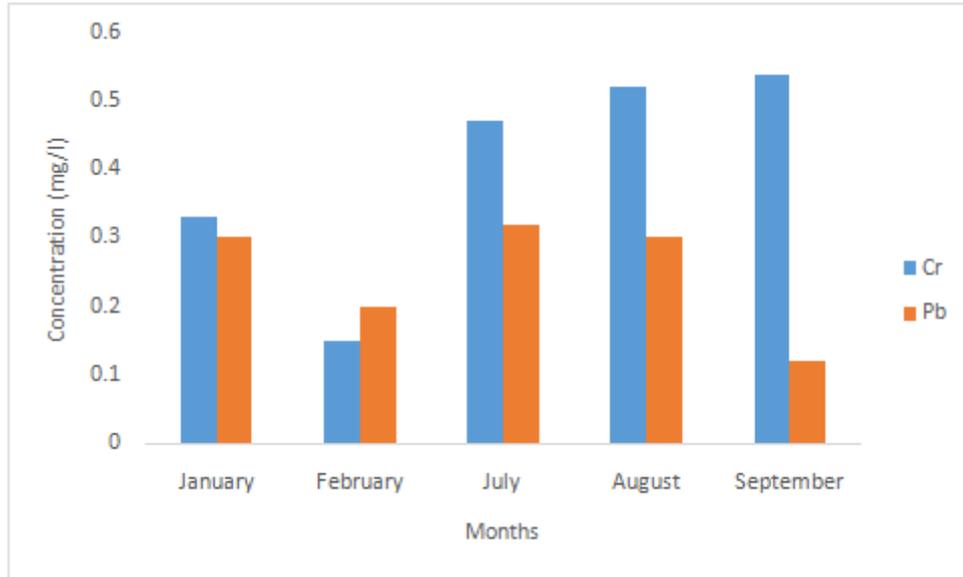


Figure.4. Mean Values of Heavy Metals in Sediment

Concentration of chromium was lowest (0.15 + 0.10mg/l) in February but highest (0.54 ± 0.14mg/l) in September while lead was lowest (0.12 ± 0.09mg/l) in September but highest (0.30 ± 0.10mg/l) in July.

3.4 Heavy Metal Accumulation in Plankton

The order of heavy metal concentrations in the plankton are chromium (0.38 ± 0.23mg/l), lead (0.21 ± 0.29mg/l). It is

therefore in the order lead < chromium (Pb< Cr). Spatially, chromium concentration in station 2 (0.22 ± 0.12mg/l) was lowest while the highest was station 3 (0.68 ± 0.30mg/l). Lead concentration was lowest (0.15 ± 0.10mg/l) in station 3 but highest (0.24 ± 0.14mg/l) in station 1 as a result of geographical variation.

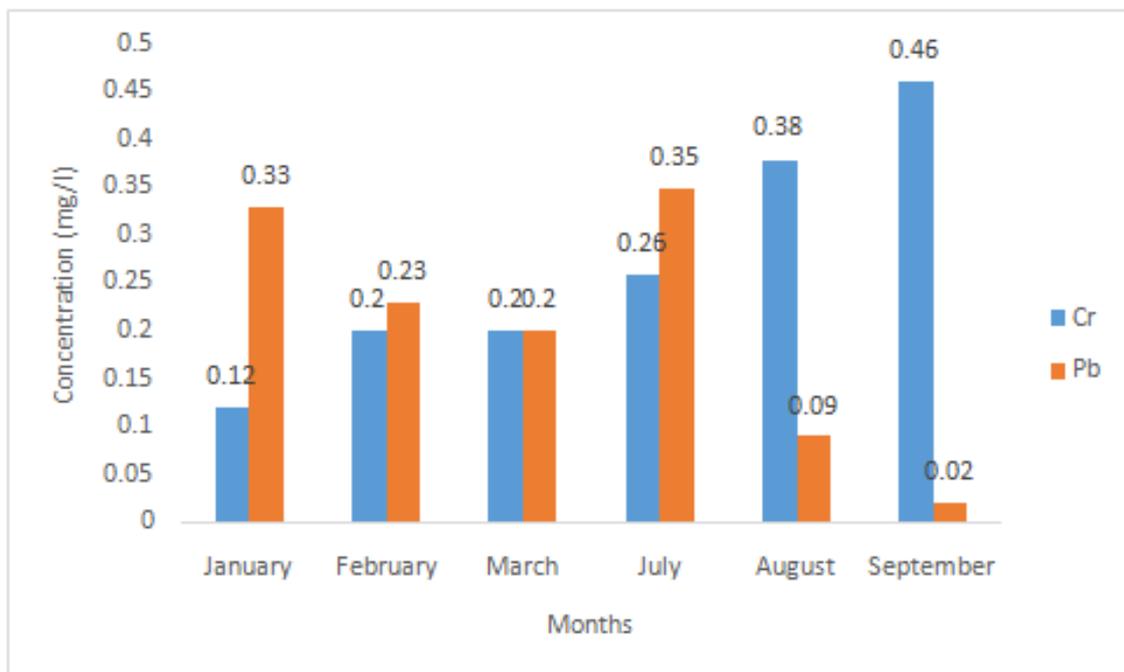


Figure.5. Mean Values of Heavy Metals in Plankton

Concentration of chromium was lowest (0.12 ± 0.01mg/l). Lead was lowest in September (0.02 ± 0.00mg/l) but highest (0.35 ± 0.10mg/l) in July.

Table .3. Spatial Mean Values of Heavy Metals in Plankton

Station	Cr	Pb
1	0.25±0.13 ^b	0.24±0.13 ^a
2	0.22±0.12 ^b	0.23±0.12 ^a
3	0.68±0.30 ^a	0.15±0.10 ^a
OverallMean	0.38±0.23	0.21±0.20

3.5 Model Validation

Table.4. Comparison between Experimental and Model results of concentration of Chromium in Water

Experiment _{crw}	Models _{crw}	Error
0.02	0.16	-0.14
0.30	0.20	0.10
0.30	0.24	0.06
0.43	0.28	0.15
0.16	0.33	-0.17
0.37	0.37	0.00

Table .5. Comparison between Experimental and Model results of concentration of Chromium in Sediment

Experiment _{crs}	Models _{crs}	Error
0.33	0.20	0.13
0.15	0.27	-0.12
0.22	0.34	-0.12
0.47	0.41	0.05
0.52	0.48	0.04
0.54	0.55	-0.01

Table .6. Comparison between Experimental and Model results of concentration of Chromium in Plankton

Experiments _{crp}	Models _{crp}	Error
0.12	0.12	0.00
0.20	0.18	0.02
0.20	0.24	-0.04
0.26	0.30	-0.04
0.38	0.36	0.02
0.46	0.40	0.06

Table .7. Comparison between Experimental and Model results of Concentration of Lead in Water

Experiments _{pbw}	Models _{pbw}	Error
0.30	0.27	0.03
0.17	0.23	-0.06
0.17	0.20	-0.03
0.30\	0.18	0.12
0.001	0.13	-0.129
0.15	0.10	0.05

Table.8. Comparison between Experimental and Model results of Concentration of Lead in Sediment

Experiments _{pbs}	Models _{pbs}	Error
0.30	0.45	-0.15
0.20	0.36	-0.16
0.13	0.27	-0.14
0.32	0.18	0.14
0.30	0.09	0.21
0.12	0.49	-0.37

Table .9. Comparison between Experimental and Model results of Concentration of Lead in Plankton

Experiments _{pbp}	Models _{pbp}	Error
0.33	0.33	0.00
0.23	0.28	-0.05
0.20	0.23	-0.03
0.35	0.17	0.18
0.09	0.12	-0.03
0.02	0.07	-0.05

4.0 CONCLUSION

This research was carried out to ascertain the plankton community of Elechi Creek in relation to physicochemical parameters and some heavy metals. Based on the findings, and results, the following facts were deduced:

1. Water temperature of the creek varied with station and season.
2. pH values were purely acidic and showed no significant difference both in season and station.
3. Electrical conductivity and dissolved oxygen did not vary with season but vary with station.
4. Biological oxygen demand, chemical oxygen demand and total hydrocarbon content showed significant difference with season and station.
5. The highest heavy metal concentration among the four investigated was chromium.
6. Heavy metal accumulation in plankton was higher than that of water and sediment.
7. Heavy metals also showed significant difference with seasons and stations.

5.0. REFERENCES

- [1]. Arruti, A., Fernandez-Olmo, I. & Irabien, A. (2010). Evaluation of the Contribution of Local Sources to Trace Metals Levels in Urban PM_{2.5} and PM₁₀ in the Cantabria Region. *Journal of Environmental Monitoring*. 12(7), 1451-1458.
- [2]. Bradl, H. (2002). Heavy Metals in the Environment: Origin, Interaction and Recommendation. London: Academic Press.
- [3]. Duffus, J. H. (2002). Heavy Metals- A Meaningless Term? *Pure and Applied Chemistry*. 74(5), 793-807.
- [4]. Ekeh, C.A. (2005). The Study of Plankton in Amadi and Nwaja Creeks of the Upper Bonny Estuary. An MSc Thesis submitted to RSUST, Port Harcourt.
- [5]. Ezra, A.G. & Nwankwo, D.I. (2001). Composition of Phytoplankton Algae in Gubi Reservoir, Bauchi, Nigeria. *Journal of Aquatic Sciences*, 16(2), 115-118.
- [6]. Ezra, J.M.D. Krom & T. N. Neuwirth (1991). Daily Oxygen Variation in Marine Fish Ponds. Elat, Israel, *Aquaculture*, 84, 89-305.
- [7]. He, Z. L., Yang, X. E. & Stoffella, P.J. (2005). Trace Elements in Agro-ecosystems and Impacts on the Environment. *Elementary Medical Biology*. 19(2-3), 125 - 140.
- [8]. Merrett, S. (2000). Industrial Effluent Policy: Economic Instruments and Environmental Regulation. *Water Policy*. 2, 201-211.
- [9]. Moss, J. C., Hardaway, C. J., Richter J. C. & Sneddon, J. (2010). Determination of Cadmium, Copper, Iron, Lead, Zinc in Crayfish [*Procambrus Clarkii*] by Inductively Coupled Plasma Optical Emission Spectrometry: A study over the 2009 Season in Southwest Louisiana. *Microchemical Journal*. 95, 5-10.
- [10]. Neal, C. & Davies, H. (2003). Water Fluxes for Eastern UK Rivers entering the North Sea: A Summary of Information from the Land Ocean Interaction Study (LOIS). *The Science of the Total Environment*. 314-316, 821-882.
- [11]. Stern, B. R. (2010). Essentiality and Toxicity in Copper Health Risk Assessment: Overview, Update and Regulatory Considerations. *Journal of Toxicology and Environmental Health, Part A*. 73 (2-3), 114-127.
- [12]. Tang, A., Liu, R., Ling, M. & Wang, J. (2010). Distribution Characteristics and Controlling Factors of Soluble Heavy Metals in the Yellow River Estuary and Adjacent Sea. *Procedia Environmental Sciences*. 2, 1193-1198.
- [13]. United States Environmental Protection Agency (USEPA). (2002). Methods for Evaluating Wetlands. Office of Water, United States Environmental Protection Agency. Washington, D.C. EPA - 822 -R -02-012.
- [14]. Vuković, Ž., Radenković, M., Stanković, S. J. & Vuković, D. (2011). Distribution and Accumulation of Heavy Metals in the Water and Sediments of the River Sava. *Journal of the Serbian Chemical Society*. 76, 795-803.
- [15]. Yan, J., He, Y., & Huang, H. (2007). Characteristics of Heavy Metals and their Evaluation in Sediments from Middle and Lower Reaches of the Huaihe River. *Journal of China University of Mining & Technology*. 17, 414-417.
- [16]. Yang, Z., Wang, Y., Shen, Z., Niu J. & Tang, Z. (2009). Distribution and Speciation of Heavy Metals in Sediments from the Mainstream, Tributaries, and Lakes of the Yangtze River Catchment of Wuhan. *China Journal of Hazardous Materials*. 166, 1186-1194.
- [17]. Zhang, C., Qiao, Q., Piper, J. D. A. & Huang, B. (2011). Assessment of Heavy Metal Pollution from Iron-Smelting Plant in Urban River Sediments Using Environmental Magnetic and Geochemical Methods. *Environmental Pollution*, 159(10), 1-14.
- [18]. Zhang, H, Cui B, Xiao R, Zhao H (2010). Heavy Metals in Water, Soils, and Plants in Riparian Wetlands in the Pearl River Estuary, South China. *Procedia Environmental Sciences*. 21, 344-1354.